

FORMULATION AND EVALUATION OF ANTIMICROBIAL GELS FOR THE TREATMENT OF PARONYCHIA

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ABSTRACT

Objective: The aim of the study was to design and develop a gel based drug delivery system containing combinational drugs (ketoconazole, neomycin sulphate and diclofenac) for the effective treatment of Paronychia.

Methods: The drugs used are ketoconazole, neomycin sulphate and diclofenac. The first two drugs provide an antifungal and antibacterial action and the last drug with a pain relieving effect. Two formulations of gels F1 and F2 were prepared using polymers like carbopol 934 and xanthan gum respectively. The amounts of drugs and other ingredients were kept as constant in both formulations. The prepared formulations were then evaluated by visual examination, pH, drug content, spreadability, extrudability, drug release study, *in vitro* antibacterial study, *in vitro* antifungal study, stability studies and *in vivo* antibacterial study.

Results: The obtained results were analyzed and compared. All the test results were within the accepted limit. The physicochemical properties of the gels were assessed and it was found that the two formulations have enough gel consistency with good spreadability and extrudability. The drug content and drug release studies of the prepared gels were done and the results showed that the all the three drugs were properly loaded into the gel system, with good drug release profile. The antimicrobial activities of the formulated gels were proved by both *in vitro* antifungal and antibacterial studies. The *in vivo* antibacterial studies revealed a significant reduction in bacterial count in wistar rats treated with prepared gel when compared with standard drug solution. From among all the developed formulations, F1 formulation with carbopol 934 has got a slight superior property when compared with formulation F2 xanthan gum as gelling agent.

Conclusion: On the basis of the evaluation studies it was concluded that the drugs (ketoconazole, neomycin sulphate and diclofenac) were successfully incorporated into the different topical gel preparations with good physicochemical properties and antimicrobial activity. Therefore, it was concluded that our formulae could be very promising topical alternative for the treatment of Paronychia.

Keywords: Paronychia, Gel, Topical formulation, Carbopol, Xanthan gum, Ketoconazole, Neomycin, Diclofenac, Antibacterial study, Drug delivery

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INTRODUCTION

Nowadays different methods like spreading, rubbing, spraying, instillation are used as various mechanisms for topical preparations; it can be applied directly to the body [1]. Topical route is one of the most effective routes of drug administration for

treating the skin disorders and they can enhance drug delivery through systemically and non-systemically [2]. The structure of nail consists of nail matrix, nail plate, nail bed, cuticle, nail folds [3] (fig. 1). The nail bed contains two portions (germinal matrix and sterile matrix), which are involved in production, migration and maintenance of nail [4].

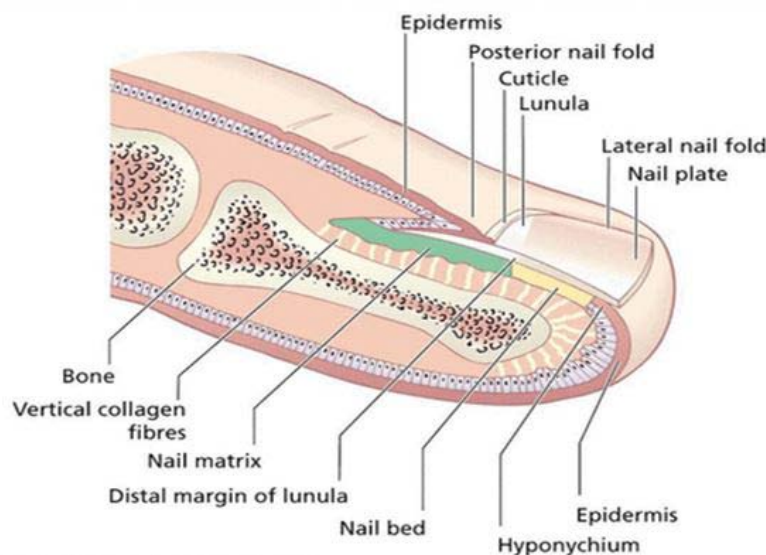


Fig. 1: Longitudinal section of nail [22]

Paronychia is a type of inflammatory disease, occurs in tissues surrounding of the nail plate. It is a most affected infection of the hand. Paronychia is mainly two types acute and chronic. Acute Paronychia is mainly seen in people who bite their nails, long nails or occurs in manicures. The disease etiology is that this type infection will break the soft tissue seal on dorsal periphery of the nail, they will enter and forming colonies after formation of abscess occurs by the offending organism. The disease starts in the lateral nail folds and it will spread other parts, *Staphylococcus aureus* is the disease causing organism [4, 5].

Chronic Paronychia is persistent type infection, disease affected mainly in the eponychium, it is somewhat different from acute Paronychia. The Paronychia is mainly seen in people they are chronically exposed to water, alkali etc. The etiology is similar like acute Paronychia. It is caused by bacterial infection; continue by super infection and colony formation of eponychium with a fungus such as *Candida albicans*. The symptoms or signs of chronic Paronychia are the characteristics of longitudinal grooves on the surface of the nail plate, followed by long term damage to the germinal tissues in the eponychium [4, 5]. The acute and chronic Paronychia affected fingers is represented in fig. 2.



Fig. 2: Acute and chronic paronychia affected fingers [4]

Surgical treatment is another approach towards Paronychia [6]. Currently there are no marketed formulations are available in the market to treat Paronychia. Gel is a semisolid system contains two phases, a gelling agent and a liquid. Topical formulations have lot of application than other routes, like they will attain local action, percutaneous penetration of the medicament or they having an emollient as well as protective action. The drug inside the gel has the ability to penetrate the skin and provide action at the site [7].

Due to the non availability of a suitable dosage form to treat Paronychia and the difficulty in assessing the causative agent (bacteria, fungus or both) in the beginning of infection has lead to

develop gel based drug delivery system containing combinational drugs for the effective treatment of Paronychia. The drugs used are ketoconazole, neomycin sulphate and diclofenac. The first two drugs provide an antifungal and antibacterial action and the last drug with a pain relieving effect. As Paronychia causes a nail piercing pain and the causative agents cannot be detected in the beginning the prepared gel system will be helpful in treating the people with Paronychia.

MATERIALS AND METHODS

Materials

Ketoconazole, neomycin sulphate and diclofenac were obtained as a gift samples from IPCA laboratories Ltd, Mumbai (India). Carbopol 934 and xanthan gum were from Sance pharmaceuticals, Kerala (India). Methyl paraben, propylene glycol and other reagents were of the Pharmacopoeial grade.

Preparation of standard graphs of ketoconazole, neomycin and diclofenac

100 mg of ketoconazole was dissolved in water and the absorbances of diluted concentrations were measured out in UV at 660 nm. The same method was followed for both neomycin and diclofenac but the absorbance was determined at 277 nm and 276 nm respectively [8, 26].

Compatibility study by FTIR

In order to study the drug compatibility in the formulations, FTIR spectrum of drugs with polymers were taken. It was done by KBr pellet method using FTIR. FTIR spectra of ketoconazole, neomycin, ketoconazole+neomycin, carbopol 934, xanthan gum, carbopol 934+ketoconazole, carbopol 934+neomycin, xanthan gum+ketoconazole and xanthan gum+neomycin were taken [8].

Solubility

Solubility study was done using different solvent system such as water, ethanol and phosphate buffer (pH 5.5 and 7.4) [9].

Formulation study

Working formula was for a quantity of 10 g of gel. Initially 0.5 g of carbopol 934 was taken in a beaker and dispersed with sufficient quantity of water using glass rod. Then it was allowed to get hydrate. After that it was neutralized with triethanolamine. Then accurate amount of drug was added with gentle stirring and the desired gel was formed [1]. In the second formulation (F2), 0.5 g of xanthan gum was replaced with 0.5 g carbopol 934. The final weight of the formulation was maintained as 10 g [10]. Working formula for formulation of gel is depicted in table 1.

Table 1: Working formula for formulation of gel

S. No.	Ingredients	F ₁	F ₂
1	Ketoconazole	0.4 g	0.4 g
2	Neomycin sulphate	0.1 g	0.1 g
3	Diclofenac	0.2 g	0.2 g
4	Carbopol 934	0.5 g	
5	Xanthan gum		0.5 g
6	Propylene glycol	4 ml	4 ml
7	Glycerin	2 ml	2 ml
8	Triethanolamine	0.06 ml	0.06 ml

Evaluation of gel

Physicochemical parameters

The prepared gels were evaluated for its physicochemical properties like clarity, colour, odour, grittiness and pH. The clarity was examined by visual examination against black or white background and the presence of gritty particles in prepared gels was tested microscopically. The prepared formulations were spread on a mounting glass plate and viewed under microscope [12]. The pH of the prepared gel formulations was determined by digital pH meter (Systronics pH meter Model 6), it was calibrated using standard buffer solutions at pH 4, 7 and 9 [11, 22].

Spreadability

It is determined by the following method. The instrument consists of a wooden block, and a pulley at one end. First of all a glass was fixed to the wooden block using an adhesive. About 2 g was gel was placed on the glass. Another slide of same dimension (with a hook) was placed over the fixed slide and the gel got sandwiched. Almost 1 kg weight was placed over the slide to remove the air entrapment. Then a weight of 30 g is attached to the hook of the slide with a string and allows some time to move the top slide through the fixed slide. Notice the time (in s) taken up by the top slide to cover a distance of 4.6 cm. A shorter interval indicates better spreadability [13].

Spreadability, $S = M(L \div T)$

S = Spreadability M = Weight in pan

L = Length moved by the glass slide T = time taken to separate the slides

Extrudability study

This method was used to analyze the force required to squeeze the content out of the tube. aluminium collapsible tube filled with 10 g gels was compressed and extrudability of the formulation was determined in terms of weight in grams required to extrude a 0.5 cm ribbon of gel in 10 s [14].

Drug content determination

The gel having a weight of 500 mg was weighed and solubilized in 50 ml of phosphate buffer solution (PBS) pH 5.5 and shakes well to extract the drug and filtered. The drug content was analysed spectrophotometrically (ketoconazole 660 nm, neomycin 277 nm, diclofenac 276 nm) [15].

In vitro drug release study

The study was done using Franz diffusion cell with cellophane membrane as barrier and 100 ml of PBS pH5.5 was used in the receptor compartment, then 1 g of gel (expected to contain 40 mg of ketoconazole, 10 mg neomycin and 20 mg of diclofenac) was spread uniformly on the membrane. The drug concentration on the receptor fluid was determined spectrophotometrically (ketoconazole 660 nm, neomycin 277 nm, diclofenac 276 nm) against appropriate blank (sampling Time 0, 5, 10, 15, 30, 60, 120 min). The experiment was carried out in triplicate [16].

In vitro antibacterial study

An agar-well diffusion method was used for determination of antibacterial activity of neomycin sulphate. The gel samples were dissolved in PBS, pH5.5. The cultured bacteria (*Staphylococcus aureus*) were suspended in sterile water and diluted to 10 colony found per unit (CFU)/ml. The suspension (100 µl) was spread onto the surface of nutrient agar medium. Wells (4.6 mm in diameter) were cut with a sterile borer and 60 µl formulations were added into them. PBS solution was used as negative control. Incubation of the inoculated plates was done at 37 °C for 24 h. The diameter of inhibition zone (DIZ) was evaluated and thereby antibacterial activity also. All tests were performed in triplicates [17, 23].

In vitro antifungal study

The antifungal activity of ketoconazole was done by agar-well diffusion method, in which the *Candida albicans* was inoculated with molten potato dextrose agar at 45 °C and allowed to set in a petri dish. Wells (4.6 mm in diameter) were cut in a similar way as for the antibacterial activity and 60 µl formulations were added into them. PBS was used to prepare the negative control. The plates were incubated at 28 °C for 3 d after which DIZ were measured [18].

Stability study

The prepared formulations (F1 and F2) were maintained at room temperature over a period of two months in three types of storage containers (plastic transparent, glass transparent and collapsible

metallic tubes). The physical appearance, homogeneity, syneresis and drug contents were evaluated after 2 mo [19]. The experiments were performed in triplicates.

Optimization of gels

The prepared gels were optimized by using the evaluated data. All the test parameters were taken into account [20].

In vivo antibacterial study

The study was conducted at central animal house, Amrita Institute of Medical Science, Kochi. All the experimental procedures were approved (IAEC/2017/3/11) and performed in accordance with the standards prescribed by the institutional animal ethical committee. 12 adult male rats of Wistar strain with body weighing 300 to 350 g, maintained under controlled light and temperature were used in this study. The animals were caged (one animal per cage) with standard food and water *ad libitum*. The animals were grouped into three, Group 1: F1 formulation Group 2: neomycin solution and Group 3: control (water for injection). *Staphylococcus aureus* bacteria were obtained from microbiology lab, Amrita School of Pharmacy and were used in this study. The anesthetized animals were placed in prone position, and it was sterilized with alcohol-iodine. A punch was used to achieve the wounds of approximately 8 mm diameter allowing the removal of a circular area of skin, on the middle portion of the medium sagittal plane. After its preparation, the wounds were colonized with a standard solution of *Staphylococcus aureus* and quantitation of bacterial population was done. Then, the 3 groups were applied with equal aliquots of F1 formulation, neomycin solution and water for injection respectively. Each group of mice receiving a particular treatment regimen was housed separately in a ventilated cage with appropriate bedding, food, and water. Mice were checked twice daily during infection and treatment to ensure no adverse reactions were observed. After 48 h, exudates were collected from the wounds for quantitation of bacterial population. The materials were processed and cultured on selective MacConkey's agar medium. The agar plates were incubated at 37 °C and examined after 24 h. Any growth in the plates of bacteria of the same biotype as cultured in the wounds was considered positive and expressed as colony forming units per gram of tissue [21].

RESULTS AND DISCUSSION

Absorbance values of ketoconazole, neomycin and diclofenac are shown in (table 2) and standard graphs were plotted between concentrations on X axis, absorbance on Y axis (fig. 3).

The FTIR analysis (fig. 4, fig. 5 and fig. 6) showed that there are no incompatibilities in the formulation. All the identification peaks of drugs are existing in the formulation also. The identification peaks of ketoconazole are C=O Stretching vibration of carbonyl group-1653 cm^{-1} , C-O stretching of aliphatic ether groups-1031.92 cm^{-1} , C-O stretching of cyclic ether-1271 cm^{-1} and chlorine group-727 cm^{-1} . The identification peaks of neomycin OH stretching-3500 cm^{-1} , N-H bending-1666 cm^{-1} and ether frequency-1180 cm^{-1} . So FTIR analysis revealed that the drug combinations are compatible and stable with other drugs and gelling agents [10, 11, 25].

Solubility

Solubility study was done using different solvent system. The obtained result was shown in (table 4)

Table 2: Absorbance values of Ketoconazole, Neomycin and Diclofenac

Concentration ketoconazole (µg/ml)	Absorbance at 660 nm	Concentration neomycin (µg/ml)	Absorbance at 277 nm	Concentration diclofenac (µg/ml)	Absorbance At 276 nm
0	0	0	0	0	0
0.25	0.142	0.10	0.006	10	0.112
0.3	0.286	0.20	0.013	20	0.223
0.35	0.431	0.30	0.019	30	0.32
0.4	0.569	0.40	0.025	40	0.441
0.45	0.723	0.50	0.033	50	0.545

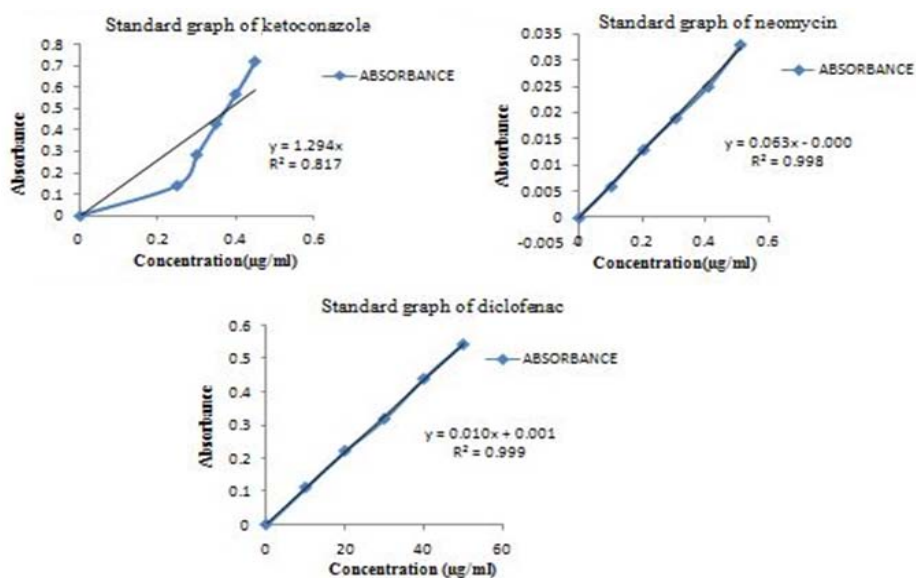


Fig. 3: Standard graphs of ketoconazole, neomycin, diclofenac compatibility study by FTIR

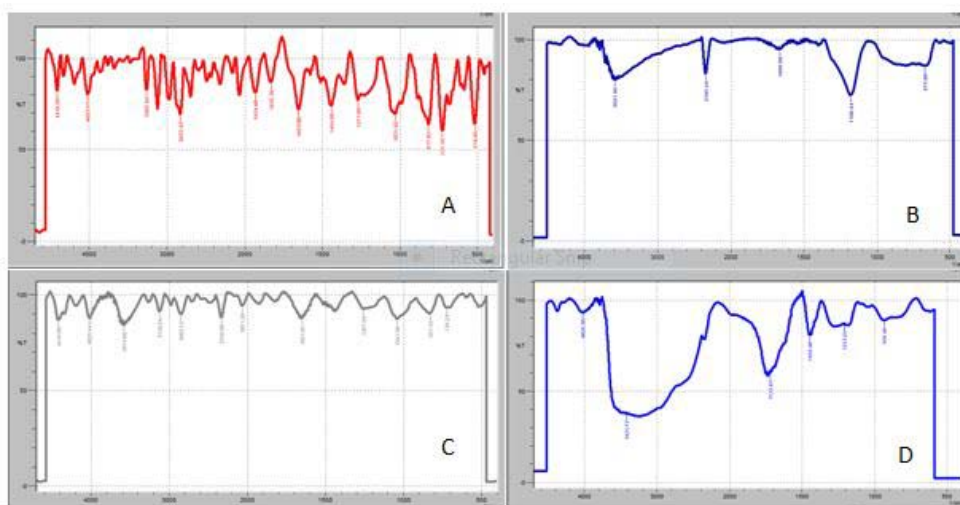


Fig. 4: FTIR spectra of A) Ketoconazole, B) Neomycin, C) Ketoconazole+neomycin and D) carbopol 934

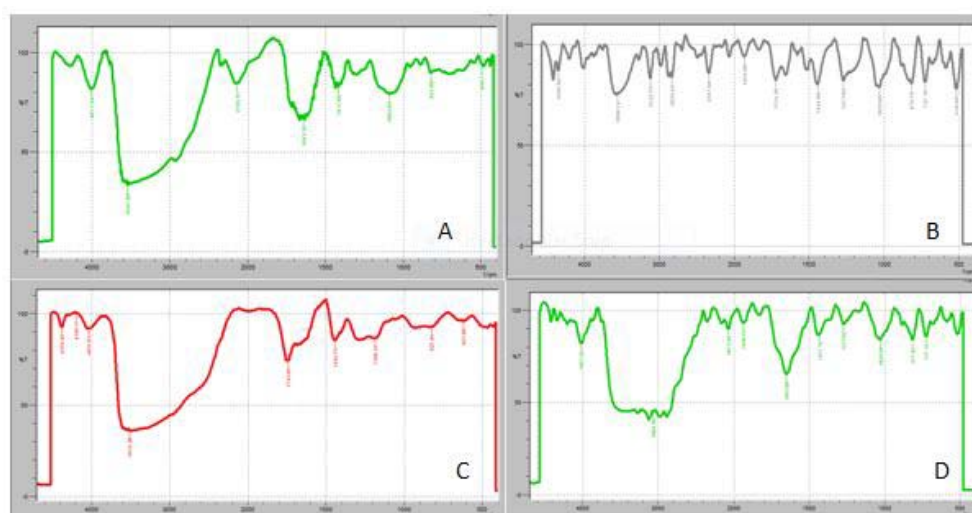


Fig. 5: FTIR spectra of A) Xanthan Gum, B) Carbopol 934+Ketoconazole, C) Carbopol 934+Neomycin and D) Xanthan Gum+Ketoconazole

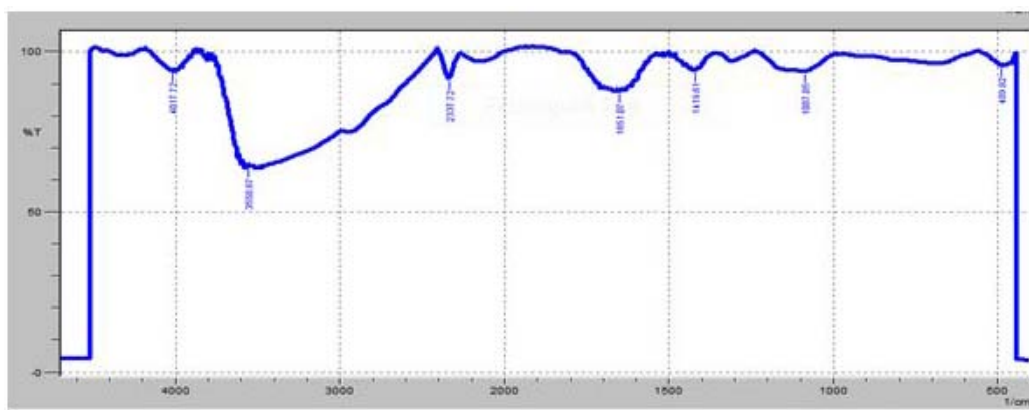


Fig. 6: FTIR spectrum of xanthan gum+neomycin

Table 3: Solubility profile of ketoconazole, neomycin and diclofenac in different solvents

Drug	Solubility in solvents		
	Water	Ethanol	Phosphate buffer (pH 5.5 and 7.4)
Ketoconazole	Soluble	Soluble	Soluble
Neomycin	Soluble	Insoluble	Soluble
Diclofenac	Soluble	Soluble	Soluble
(n=3)			



Fig. 7: The prepared gels F1 (carbopol 934) and F2 (xanthan gum) as gelling agents

Formulation study

The formulations of gels were prepared as prescribed method and it is shown in (fig. 7).

Physicochemical parameters

The prepared formulations were evaluated for clarity test and the study results show that, both the gels were clear and free of particles. The formulations exhibited a good homogeneity. The gel with carbopol 934 as gelling agent showed a good transparency when compared with the gel with xanthan gum as gelling agent. Both the formulations were viscous in nature. Xanthan gum

showed a translucent nature. The formulated gels were visually evaluated and it was found to be clear and white cloudy appearance for gel with carbopol 934 as gelling agent whereas the other formulation appeared as slight yellowish color with good clarity. Both the formulations have got a rose fragrance. The samples were free of particulate matter was seen under light microscope. Hence the gel is free of particular matter and can be used for any topical preparation. The pH of the formulated gels was found to be 5.7 ± 0.1 for carbopol 934 gel and 6.4 ± 0.1 for xanthan gum gel. The obtained pH resembles to skin pH and compatible for topical formulation [10, 12, 15]. Physicochemical properties of the gels are summarized in table 4.

Table 4: Physicochemical properties of the gels

Topical gels	Color	Homogeneity	pH	Odour
F ₁	Transparent	Homogenous	5.7 ± 0.1	Rose fragrance
F ₂	Translucent yellowish	Homogenous	6.4 ± 0.1	Rose fragrance

Data expressed as mean \pm SD (n=3)

Spreadability and extrudability study

The spreadability test was performed as prescribed in the procedure. The test results revealed that the gel with carbopol 934 as gelling agent has got a higher spreadability when compared with xanthan gum gel formulation. The values of spreadability were 4.6 cm and 4.2 cm respectively for both carbopol 934 and xanthan gum gels [18, 20]. The slight change in spreadability may be due to the change in polymer. All the two formulations showed a good extrudability. There is no significance change in the extrudabilities of F1 and F2. So, plug flow is limited in this

formulation and is easy to squeeze out the formulation from the collapsible tube and there is a uniformity of drug contents in the extruded mass [20].

Drug content determination

Results of drug content are shown in (table 5). After the formulation of the two types of gels, the drug content of the formulated gel was estimated and the results were in the official limits. The drug content determination also showed the uniformity of drug distribution in the gel [18, 20].

Table 5: Drug content of the gels

Topical gels	Ketoconazole	Neomycin	Diclofenac
	%W/W	% W/W	% W/W
F ₁	98.4±0.7	97.4±1.1	99.2±0.8
F ₂	98.0±1.0	96.0±0.9	98.6±1.0

*Data expressed as mean±SD (n=3)

In vitro drug release study

It was observed that the release of the drugs from its different formulae didn't show any significant difference in drug release. On comparison of the obtained *in vitro* data; it was found that F1 showed a bit faster release of drugs when compared with F2. The amount of the drug released after 2 h were 40.1±1.22%, 40.1±1.24%, and 53±1.24% for ketoconazole, neomycin and

diclofenac respectively from F1, whereas in F2 the release was 38±1.27%, 38±1.20% and 52±1.27% respectively.

On analysis, it was found that almost 30% of the drug is been released within 5 min. So it is an indication that the gel will act very fast in the infected area. Especially the drug release for diclofenac was almost 40% within 5 min and may get a faster pain relieving action in the affected area. It is shown in (table 6).

Table 6: In vitro drug release profile

Time (min)	F1			F2		
	K1	N1	D1	K2	N2	D2
0	0	0	0	0	0	0
5	31.0±1.25	30.0±1.22	41.5±1.21	29.0±1.22	28.0±1.24	38.0±1.20
10	32.2±1.20	32.5±1.25	42.5±1.22	30.2±1.25	29.2±1.25	39.2±1.22
15	33.4±1.21	33.7±1.20	45.5±1.20	31.4±1.28	30.4±1.22	40.4±1.24
30	35.0±1.22	36.5±1.27	48.5±1.25	33.9±1.24	32.4±1.24	44.0±1.28
60	38.5±1.20	38.0±1.22	51.0±1.24	36.0±1.27	35.5±1.27	48.0±1.25
120	40.1±1.22	40.1±1.24	53.0±1.24	38.0±1.27	38.0±1.20	52.0±1.27

*Data expressed as mean±SD (n=3) K-ketoconazole, N-neomycin and D-diclofenac

In vitro antibacterial and antifungal study

The antibacterial activities of neomycin from its different gel formulae were compared. The results were satisfactory. In this case the DIZ was same (33±1 mm) for both F1 and F2. The results of all formulae were satisfactory for antifungal study also. A slight increased activity was observed with F1 where the inhibition zone reaches 37±1 mm whereas F2 where the inhibition was 35±1 mm [17, 18]. There is not much significant difference in antifungal activity of ketoconazole in both the formulations.

Stability study

On evaluation of the formulations, it was found that there is some slight change in the physical characters of the formulations (F1 and F2) that stored in plastic container when compared with other containers. It is shown in (table 7). The formulations stored in

plastic containers showed a red coloration; it may be due to the leaching of the container. The formulations in other containers were found to be stable with no coloration. Other physical parameters were same for all the formulations during the entire storage period. There were no syneresis and grittiness in all the stored formulations [19]. The drug content results revealed that there is no significant change in drug content after stability study.

Optimization of gel

The optimization of the prepared gels was done by comparing the results of different evaluation tests. By comparing all the data it was found that the F1 formulation with carbopol 934 as gelling has got a slight superior property when compared with F2; where xanthan gum was used as gelling agent. The change in evaluation test results may be mainly due to the change in polymer.

Table 7: Stability data of F1 and F2 formulations after two months (K-ketoconazole)

Formulation	Container	Physical Parameters				Syneresis	Drug content, K
		Color	Homogeneity	Odour	Grittiness		
F1	Plastic	Red	Homogeneous	Rose	No	No	98.0±0.1%
F2	Plastic	Red	Homogeneous	Rose	No	No	96.5±0.5%
F1	Glass	White transparent	Homogeneous	Rose	No	No	98.1±0.8%
F2	Glass	Yellowish translucent	Homogeneous	Rose	No	No	97.2±0.5%
F1	C. Tube	White transparent	Homogeneous	Rose	No	No	98.5±0.4%
F2	C. Tube	Yellowish translucent	Homogeneous	Rose	No	No	98±0.6%

*Data expressed as mean±SD (n=3), K-Ketoconazole

In vivo antibacterial study

The change in colony forming units for each group was investigated and expressed as mean values. Comparison of the values of CFU between the start and end times for each group were calculated (table 8). The values were calculated in terms of % variation in CFU in each group also. From the results, it was found that there is so much reduction in the CFU in the case of group 1 (92.38%) and group 2 (95.76%), which were treated with F1 formulation and

neomycin solution respectively whereas in Group 3, a sharp increase in CFU was found. At the same time there is no significant difference in reduction in the CFU between Group 1 and 2. However in group 3 treated with distilled water showed a 68% increment in CFU. The study reveals that the Formulation 1 has significant antibacterial action in reducing the *Staphylococcus aureus* and can be used in the treatment of Paronychia. In accordance with the presented results, it can conclude that Formulation 1 can be an alternative for current treatment and surgical method [21, 23].

Table 8: Comparison of the values of CFU between the start and end times for each group

Animal number	Group 1		Group 2		Group 3	
	Start	End	Start	End	Start	End
1	2555	230	2600	108	2503	4800
2	2203	200	3033	55	2700	4900
3	2400	150	2879	230	2400	3300
4	2144	130	3100	100	2452	3900
Mean	2325.5±118.1	177.5±45.7	2903±222.1	123.5±74.8	2513.8±131.0	4225±763.2

*Data expressed as mean±SD (n=4)

CONCLUSION

On the basis of the evaluation studies we can conclude that the drugs (ketoconazole, neomycin and diclofenac) were successfully incorporated into the different topical gel preparations. From among all the developed formulation, the formula F1 (carbopol 934 as gelling agent) showed comparatively good spreadability, extrudability, drug content, drug release and antimicrobial effect. But there is not much significant difference in the evaluated parameters of both formulations. The antifungal and antibacterial effects of these formulations also found to be satisfactory. Therefore, it was concluded that our formulae could be very promising topical alternative for the treatment of Paronychia. However, further preclinical and clinical studies are required to prove its application in human.

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AUTHORS CONTRIBUTIONS

All the author have contributed equally

CONFLICTS OF INTERESTS

Authors have no conflict of interest

REFERENCES

- Doaa AH, Dalia A, Sally A, Abdel H, Mohamed AE. Formulation and evaluation of fluconazole topical gel. *Int J Pharm Pharm Sci* 2012;4:176-83.
- Vineet R, Khushbu G, Shikha B, Vijay KG. Management of chronic paronychia. *Indian J Dermatol* 2014;59:1-14.
- Bendar MS, Lane LB. Epionychial marsupialization and nail removal for surgical treatment of chronic paronychia. *J Hand Surg Am* 1991;16:314-7.
- Dimitris R, Geogrg L, Stamatis G. Acute and chronic paronychia. *Am Fam Physician* 2008;77:339-46.
- Keyser JJ, Eaton EG. Surgical cure of chronic paronychia by epionychial marsupialization. *Plast Reconstr Surg* 1976;58:66-70.
- Abrams RA, Botte MJ. Hand infections: treatment recommendations for specific types. *J Am Acad Orthop Surg* 1996;4:219-30.
- Kumar R, Katare OP. Lecithin organogels as potential phospholipid structured system for topical drug delivery. *AAPS PharmSciTech* 2005;6:298-310.
- V Rajpal, DP S Kohli. *Herbal drug industry*. 1st ed. New Delhi: Business Horizons; 2009.
- Vinod MT, Bharath WT, Bharat RP. Formulation and characterization of ketoprofen topical gel. *Int J Curr Pharm Clin Res* 2014;4:35-42.
- Tanwar YS. Formulation and evaluation of topical diclofenac sodium gel using different gelling agent. *AJPRHC* 2011;4:1-6.
- Japan P, Brijesh P, Hardeepsingh B, Kaushal P, Manish P. Formulation and evaluation of topical aceclofenac gel using different gelling agents. *Int J Drug Dev Res* 2011;4:23-9.
- Subrahmanyam CVS. *Text book of physical pharmaceutics*. 2nd ed. New Delhi: Vallabh Parakashan; 2009.
- Saad SD, Mir NA, Hajera T, Mazharuddin K. Studies on antibacterial and antifungal activity of pomegranate (*Punica granatum* L.). *American-Eurasian J Agric Environ Sci* 2010;9:273-81.
- Kumar TG, Kaur LP. Formulation and evaluation of topical gel of aceclofenac. *J Drug Delivery Ther* 2013;3:51-3.
- Bharat P, Atul K, Ajay C. Formulation and evaluation of gel containing miconazole nitrate an antifungal agent. *Int J Pharma Res Rev* 2013;2:18-28.
- Marwa HS, Ghada FM. Evaluation of topical gel bases formulated with various essential oils for antibacterial activity against methicillin resistant *Staphylococcus Aureus*. *Trop J Pharm Res* 2013;12:877-84.
- Susan H. Comparison of *in vitro* dialysis release methods of loperamide encapsulated liposomal gel for topical drug delivery. *Int J Nanomed* 2014;9:735-44.
- Enkelegda G, Entela H, Skerdilaid X, Ledjan M. Formulation and *in vitro* evaluation of diclofenac sodium gel. *Int J Pharm Pharm Sci* 2014;6:12-7.
- Safeela N, Fatima Q. UV spectrophotometric assay of diclofenac sodium available brands. *J Innovation Pharma Bio Sci* 2011;3:14-7.
- Ajinkya MB, Manjusha ND. Formulation and evaluation of herbal antimicrobial gel containing *Musa acumulata* leaves extract. *J Pharmacogn Phytochem* 2016;5:1-3.
- Paulo CC, Filipe AR, Ana CGB, Baldemor AKS, Daniel MP, Nadia CB, Ana PCM, Geisa PO. Proyecto academico sin fines de lucro, desarrollado bajo la iniciativa de acceso abierto. *Conscientiae Saude* 2008;7:423-9.
- Samantha E. Fungal nail infection: diagnosis and management. *Br Med J* 2009;18:26-30.
- Nina IH, Marline N, Urip H. Formulation and evaluation of herbal antibacterial gel containing ethanolic extract of mikania micrantha kunth leaves. *Asian J Pharm Clin Res* 2018;11:429-31.
- Lakshmi PK, Marka K, Aishwarya S, Shyamala B. Formulation and evaluation of ibuprofen topical gel: a novel approach for penetration enhancement. *Int J Appl Pharm* 2011;3:25-30.
- Nidhi C, Chakraborty GS. Formulation and evaluation of herbal hydro gel from *Hibiscus rosa-sinensis*. *Int J Curr Pharm Res* 2014;6:14-6.
- Amita HP, Riddhi MD. Formulation and evaluation of Sustained Release In situ ophthalmic gel of neomycin sulphate. *Bull Pharm Res* 2015;5:1-5.